$Supplementary\ Information\ for$ Efficient variance components analysis across millions of genomes

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1 Supplementary Notes

Computing the Standard Errors of the RHE-mc estimates 1.1

We obtain standard errors for RHE-mc using a block jackknife [1]. A jackknife subsample is created by leaving out a subset of observations from a dataset. The jackknife estimate of a parameter can be found by estimating the parameter for each subsample omitting the i-th jackknife block. A naive way to compute jackknife estimate requires computing the estimator of the parameters for every sub-sample. For instance, in our problem, if we define J jackknife blocks, then we need to run RHE-mc for every sub-sample which takes $\mathcal{O}(J(\frac{NMB}{\max(\log_3(N),\log_3(M))} + K^2(K+NB)))$. We propose an efficient way to compute the jackknife estimate in time $\mathcal{O}(\frac{NMB}{\max(\log_3(N),\log_3(M))} + JK^2(K+NB))$. Let \boldsymbol{X} be a $N \times M$ matrix of standardized genotypes where N and M are the number of

individuals and SNPs respectively. To generate J jackknife subsamples, we partition X into Jnon-overlapping blocks $X^{(1)}, \ldots, X^{(J)}$ such that $X = [X^{(1)}, X^{(2)}, \ldots, X^{(J)}]$. Note that for every $j, \mathbf{X}^{(j)}$ is a $N \times M_j$ matrix where M_j is the number of SNPs in the j-th block.

We create the j-th jackknife subsample by removing the j-th block $X^{(j)}$ from X. To estimate the variance components of the j-th jackknife subsample, we need to compute the corresponding quantities of the jth subsample in the normal equations (Methods). Let $K_k^{(-j)}$ be the GRM of the k-th partition which is created by removing the j-th block $X^{(j)}$ from X where $k \in \{1, \ldots, K\}$, $j \in \{1, \ldots, J\}$. In Algorithm 1.3, we show how we can compute $tr(\widehat{K_k^{(-j)}K_l^{(-j)}})$ and $y^TK_i^{(-j)}y$, for all $k, l \in \{1, \dots, K\}, j \in \{1, \dots, J\}$ efficiently.

1.2Including covariates in RHE-mc

We can extend the LMM to include covariates as follows:

$$y|\epsilon, \beta_1, \dots, \beta_k = W\alpha + \sum_k X_k \beta_k + \epsilon$$
 (1)

Here W is a $N \times C$ matrix of covariates while α is a C-vector of fixed effects. It is easy to see that the matrix $V = I_N - W(W^TW)^{-1}W^T$ is symmetric and idempotent $(\mathbf{V}^2 = \mathbf{V})$ of rank N - C. Therefore, we consider the eigendecomposition of $\mathbf{V} = \mathbf{E} \mathbf{D} \mathbf{E}^T$, where D is a diagonal matrix with N-C ones and C zeros on the diagonal (we can assume that first N-C elements are one). Now let the matrix $U_{N\times (N-C)}$ represent the first N-C columns of E. It is not hard to see that U satisfies $U^TU = I_{N-C}$, $UU^T = V$, $U^TW = 0$. Now we multiplying by U^T on both sides of the above equation:

$$U^{T}y = U^{T} \sum_{k} X_{k} \beta_{k} + U^{T} \epsilon$$
 (2)

$$cov(\boldsymbol{U}^{T}\boldsymbol{y}) = E[\boldsymbol{U}^{T}\boldsymbol{y}(\boldsymbol{U}^{T}\boldsymbol{y})^{T}] - E[\boldsymbol{U}^{T}\boldsymbol{y}]E[\boldsymbol{U}^{T}\boldsymbol{y}]$$
(3)

The matrix U^T is constant and the vector y is random. Therefore, we have $E[U^Ty] = U^TE[y]$.

$$\boldsymbol{U}^{T}\boldsymbol{y}(\boldsymbol{U}^{T}\boldsymbol{y})^{T} = (\boldsymbol{U}^{T}\sum_{k}\boldsymbol{X}_{k}\boldsymbol{\beta}_{k} + \boldsymbol{U}^{T}\boldsymbol{\epsilon})(\boldsymbol{U}^{T}\sum_{k}\boldsymbol{X}_{k}\boldsymbol{\beta}_{k} + \boldsymbol{U}^{T}\boldsymbol{\epsilon})^{T} =$$
(4)

$$\sum_i \sum_j \boldsymbol{U}^T \boldsymbol{X}_i \boldsymbol{\beta}_i (\boldsymbol{U}^T \boldsymbol{X}_j \boldsymbol{\beta}_j)^T + (\boldsymbol{U}^T \boldsymbol{\epsilon}) \sum_i (\boldsymbol{U}^T \boldsymbol{X}_i \boldsymbol{\beta}_i)^T + \sum_i \boldsymbol{U}^T \boldsymbol{X}_i \boldsymbol{\beta}_i (\boldsymbol{U}^T \boldsymbol{\epsilon})^T + \boldsymbol{U}^T \boldsymbol{\epsilon} (\boldsymbol{U}^T \boldsymbol{\epsilon})^T$$

Hence

$$E[\boldsymbol{U}^{T}\boldsymbol{y}(\boldsymbol{U}^{T}\boldsymbol{y})^{T}] = \sum_{k} \frac{\sigma_{g_{k}}^{2}}{M_{k}} (\boldsymbol{U}^{T}\boldsymbol{X}_{k}) (\boldsymbol{U}^{T}\boldsymbol{X}_{k})^{T} + \sigma_{\epsilon}^{2} \boldsymbol{U}^{T} \boldsymbol{U}$$
(5)

Using $\boldsymbol{K}_k = \frac{\boldsymbol{X}_k \boldsymbol{X}_k^T}{M_k}$, we have:

$$cov(\boldsymbol{U}^{T}\boldsymbol{y}) = \boldsymbol{U}^{T}(\sum_{k} \sigma_{g_{k}}^{2} \boldsymbol{K}_{k}) \boldsymbol{U} + \sigma_{\epsilon}^{2} \boldsymbol{I}_{N-C}$$
(6)

The MoM estimator is obtained by solving the following ordinary least squares problem:

$$(\tilde{\sigma_1^2}, \dots, \tilde{\sigma_K^2}, \tilde{\sigma_e^2}) = argmin_{(\sigma_1^2, \dots, \sigma_K^2, \sigma_e^2)} || \boldsymbol{U}^T \boldsymbol{y} (\boldsymbol{U}^T \boldsymbol{y})^T - \boldsymbol{U}^T (\sum_k \sigma_k^2 \boldsymbol{K}_k) \boldsymbol{U} - \sigma_\epsilon^2 \boldsymbol{I}_{N-C} ||_F^2$$
 (7)

We need to solve the following normal equations to estimate the variance components.

$$\begin{bmatrix} \mathbf{T} & \mathbf{b} \\ \mathbf{b}^T & N - C \end{bmatrix} \begin{bmatrix} \sigma_1^2 \\ \vdots \\ \sigma_k^2 \\ \sigma_e^2 \end{bmatrix} = \begin{bmatrix} \mathbf{c} \\ \mathbf{y}^T \mathbf{V} \mathbf{y} \end{bmatrix}$$
(8)

Here $\mathbf{V} = \mathbf{I}_N - \mathbf{W}(\mathbf{W}^T \mathbf{W})^{-1} \mathbf{W}^T$ and \mathbf{T} is a $K \times K$ matrix where $T_{k,l} = tr(\mathbf{K}_k \mathbf{V} \mathbf{K}_l \mathbf{V})$, and b is a K-vector where $b_k = tr(\mathbf{V} \mathbf{K}_k)$, and \mathbf{c} is a K-vector where $c_k = \mathbf{y}^T \mathbf{V} \mathbf{K}_k \mathbf{V} \mathbf{y}$. Commonly, the number of covariates C is small (tens to hundreds) so that including covariates does not significantly affect the computational cost. The cost of computing the elements of the normal equations 8 includes the cost of inverting $\mathbf{W}^T \mathbf{W}$ which is a $C \times C$ matrix and multiplying \mathbf{W} by a real-valued N-vector which can be done in $\mathcal{O}(C^3 + NC)$.

1.3 Streaming version

Here we describe the streaming version of RHE-mc algorithm. In Methods section, we showed that our MoM estimator satisfy the following normal equation.

$$\begin{bmatrix} \boldsymbol{T} & \boldsymbol{b} \\ \boldsymbol{b}^T & N \end{bmatrix} \begin{bmatrix} \tilde{\boldsymbol{\sigma}_g^2} \\ \tilde{\boldsymbol{\sigma}_e^2} \end{bmatrix} = \begin{bmatrix} \boldsymbol{c} \\ \boldsymbol{y}^T \boldsymbol{y} \end{bmatrix}$$
(9)

Here
$$\tilde{\sigma}_{g}^{2} = \begin{bmatrix} \tilde{\sigma}_{1}^{2} \\ \vdots \\ \tilde{\sigma}_{K}^{2} \end{bmatrix}$$
, T is a $K \times K$ matrix with entries $T_{k,l} = tr(K_{k}K_{l}), k, l \in \{1, \dots, K\}$, b is a

K-vector with entries $b_k = tr(\mathbf{K}_k) = N$ (because \mathbf{X}_k s is standardized), and \mathbf{c} is a K-vector with entries $c_k = \mathbf{y}^T \mathbf{K}_k \mathbf{y}$. Here we estimate $T_{k,l}$ as follows:

$$T_{k,l} = tr(\boldsymbol{K}_k \boldsymbol{K}_l) \approx \widehat{T_{k,l}} = \frac{1}{B} \frac{1}{M_k M_l} \sum_{k} \boldsymbol{z}_b^T \boldsymbol{X}_k \boldsymbol{X}_k^T \boldsymbol{X}_l \boldsymbol{X}_l^T \boldsymbol{z}_b$$
(10)

Here z_1, \ldots, z_B are B independent random vectors with zero mean and covariance I_N .

We read genotype matrix \boldsymbol{X}_k for every $k \in \{1, \dots, K\}$ block by block. We define J blocks over \boldsymbol{X}_k by partitioning the columns of \boldsymbol{X}_k to J groups such that $\boldsymbol{X}_k = [\boldsymbol{X}_k^{(1)} \dots \boldsymbol{X}_k^{(J)}]$.

Algorithm 1: Streaming version of RHE-mc

```
for every genotype matrix k \in \{1,..,K\} do
     for every block j \in \{1, ..., J\} do
          Read \boldsymbol{X}_{k}^{j} for every random vector b \in \{1, .., B\} do \begin{vmatrix} Z_{(k,j,b)} = \boldsymbol{X}_{k}^{j} \boldsymbol{X}_{k}^{j}^{T} z_{b} \\ \text{end} \end{vmatrix}
          v = \boldsymbol{X}_k^{jT} y \\ H_{(k,j)} = v^T v
           Release the memory allocated to X_k^j
     end
end
Let U_{k,b,0} = \sum_{j} Z_{(k,j,b)}, and
Let U_{k,b,j} = U_{k,b,0} - Z_{(k,j,b)}, for every k, b, j.
Let V_{k,0} = \sum_{j} H_{k,j},
Let V_{k,j} = V_{k,0} - H_{k,j} for every k, j
for every j \in \{0, 1, ..., J\} do
     for every pairs of genotype matrices k and l \in \{1, ..., K\} do
          \widehat{T_{k,l}} = \frac{1}{B} \frac{1}{M_k M_l} \sum_b U_{(k,j,b)}^T U_{(l,j,b)}
      end
     for every genotype matrix k \in \{1, .., K\} do
          c_k = \frac{1}{M_k} V_{(k,j)}^T V_{(k,j)}
     Solve the normal equation for j^{th} sub-sample (j=0 corresponds to the original genotype
       matrix used for computing the point estimates)
end
```

Compute the jackknife SE from the point estimates of J sub-samples.

In the above algorithm, the 3-D matrices Z and U need $\mathcal{O}(JKBN)$ memory, the 2-D matrices V and H need $\mathcal{O}(JN)$ memory. So the total space complexity will be $\mathcal{O}(JKBN)$. The total running time of this implementation is $\mathcal{O}(\frac{NMB}{\max(\log_3(N),\log_3(M))} + JK^2(K+NB)))$. For simplicity we assume that the streaming blocks are same as jackknife blocks. However, we can set the size of the streaming blocks to be different from the jackknife blocks to make the algorithm more efficient in terms of memory usage.

1.4 Parameter settings for summary statistics methods

For running LDSC we computed the LD score of each SNP within 2-Mb windows centered on the SNP. We ran LD score regression with an unconstrained intercept and with regression weights that account for correlations between association statistics at SNPs in LD and heteroscedasticity [2]. For preventing the LDSC software from dropping high-effect SNPs we used the following flags—not-M-5-50 and—chisq-max 99999.

In simulations, we ran S-LDSC with 10 binary MAF bins which are defined such that each bin contains 10% of the typed SNPs; this is done to reflect the 10 MAF bin annotations in the S-LDSC baseline-LD model [3] (see Table 5 for the details of MAF bins). In analyzing the 22 real complex traits, we run S-LDSC with baseline-LD model [3].

To run SumHer, first we computed the default LDAK weights using in-sample LD [4]. After that

we computed LD tagging using 1-Mb windows centered on each SNP and setting $\alpha = -0.25$ as recommended [5]. We used default values for the other parameter settings for running SumHer.

To do a direct comparison among LDSC, S-LDSC, and SumHer, we ran an in-sample LD version of each method meaning that we used same set of SNPs to compute LD scores and LDAK weights, perform the regression, and estimate SNP-heritability.

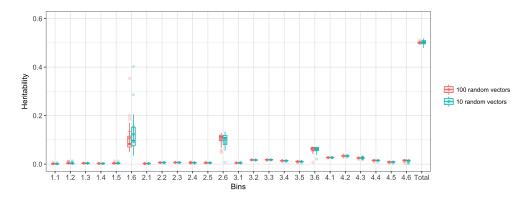
1.5 Continuous annotations

We assessed the accuracy of RHE-mc in estimating variance components with continuous annotation. We simulated a phenotype with true heritability 0.5 from 9K individuals and 15k SNPs under the GCTA model. We ran RHE-mc with single component, no annotations, and standardized genotypes. We next ran RHE-mc with single component, non-standardized genotypes, where we added a continuous annotation defined as 1/var(i) for SNP i where var(i) is the variance of SNP i across individuals. We obtain concordant estimate of genome-wide SNP heritability 0.45 ± 0.03 in the first case and 0.46 ± 0.03 in the second case.

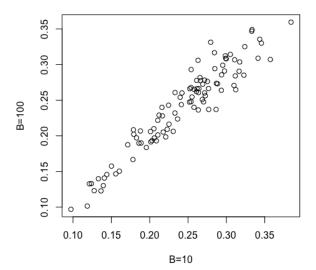
1.6 Power as a function of annotation size

To quantify the power of RHE-mc as a function of the size of an annotation, we performed simulations using N=291,273 unrelated white British individuals and M=459,792 common SNPs. We defined 8 annotations (4 MAF bins and 2 LD bins) in which we fixed the heritability of a selected bin and varied the proportion of SNPs in the selected category. We then plotted probability of rejection; the results are displayed in Supplementary Figure 11 . Furthermore, we simulated phenotypes in which we fixed the enrichment of a selected bins and varied the size of the selected bin, the results are displayed in Supplementary Table 6.

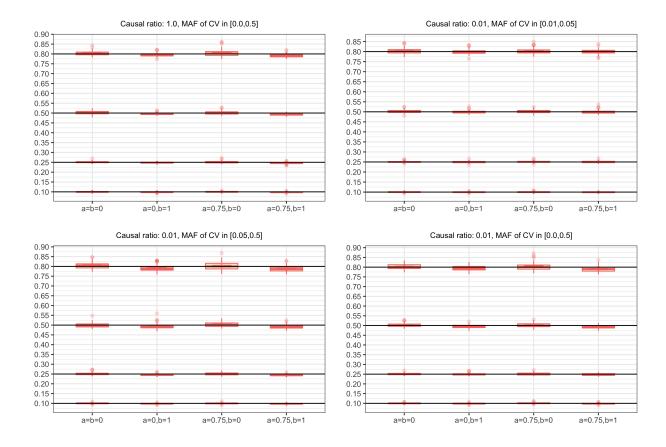
2 Supplementary Figures



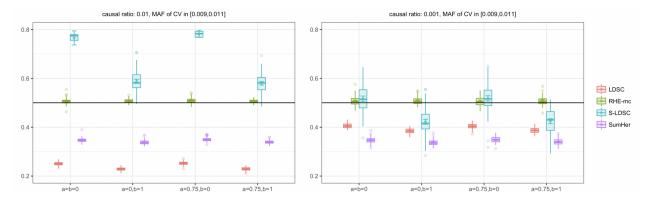
Supplementary Figure 1: Comparison of RHE-mc heritability estimates with B=10 and B=100 random vectors on large-scale simulated data (M=590K array SNPs and N=337K individuals): We ran RHE-mc with 24 bins (based on 6 MAF bins and 4 LDAK bins, see Methods). Here x-axis represents the bins (i.j denotes the bin defined based on i-th ldak bin and j-th MAF bin) and y-axis represents the heritability. Boxplot whiskers extend to the minimum and maximum estimates located within $1.5 \times$ interquartile range (IQR) from the first and third quartiles, respectively. Each box plot represents estimates from 100 simulations. Diamond points and error bars represent the mean and ± 2 SE centered on estimated heritability respectively. Mean and standard errors (SE's) are computed from 100 replicates.



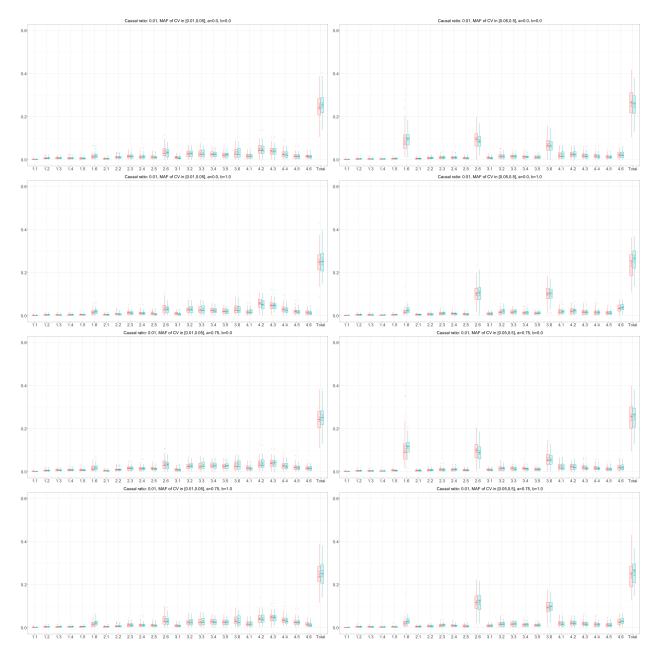
Supplementary Figure 2: Comparison of RHE-mc estimates with B=10 and B=100 on small scale data (M=590K array SNPs and N=10k individuals): We simulated 100 phenotypes such that the true total heritability is 0.25. Here x-axis represents the RHE-mc estimates when B=10, and y-axis represents RHE-mc estimates when B=100.



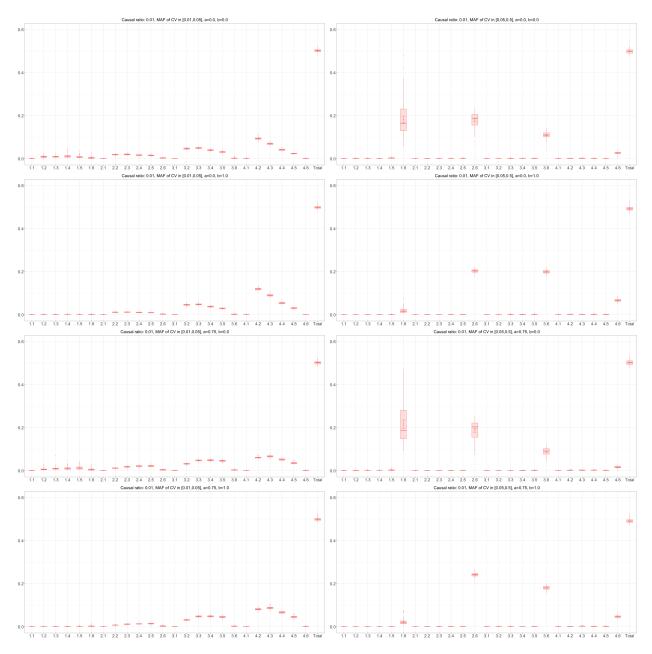
Supplementary Figure 3: Accuracy of genome-wide SNP heritability estimated by RHE-mc across 64 distinct MAF- and LD-dependent architectures in genome-wide simulations (N=337,205 unrelated individuals, M=593,300 array SNPs). For simulating the phenotypes, we chose true heritability from $\{0.1,0.25,0.5,0.8\}$, varied the ratio of causal variants (causal ratio $\in \{0.01,1.0\}$), varied the MAF range of causal variants (MAF of CV), the coupling of MAF with effect size (a=0 indicates no coupling of MAF and a=0.75 indicates coupling of MAF), and the effect of local LD on effect size (b=0 indicates no LDAK weights and b=1 indicates LDAK weights) (see Methods). We ran RHE-mc using 24 bins formed by the combination of 6 bins based on MAF as well as 4 bins based on quartiles of the LDAK score of a SNP (see Methods). Boxplot whiskers extend to the minimum and maximum estimates located within $1.5\times$ interquartile range (IQR) from the first and third quartiles, respectively. Each box plot represents RHE-mc estimates from 100 simulations.



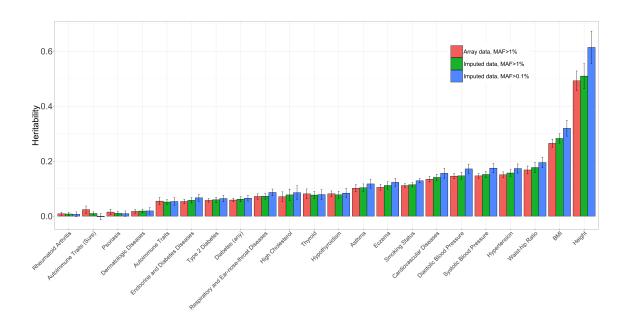
Supplementary Figure 4: Comparison of estimates of genome-wide SNP heritability from RHE-mc with LDSC, S-LDSC, and SumHer when only rare variants are causal in large-scale simulations (N = 337, 205 unrelated individuals, M = 593, 300 array SNPs). We compared methods for heritability estimation under different genetic architectures when only rare variants are causal. We set true heritability to 0.5, the MAF range of causal variants (MAF of CV) to be between [0.009, 0.011] and varied the coupling of MAF with effect size (a = 0 indicates no coupling of MAF and a = 0.75 indicates coupling of MAF), and the effect of local LD on effect size (b = 0 indicates no LDAK weights and b = 1 indicates LDAK weights) (see Methods). Here, we run RHE-mc using 24 bins formed by the combination of 6 bins based on MAF as well as 4 bins based on quartiles of the LDAK score of a SNP (see Methods). We run S-LDSC with 10 MAF bins (see Supplementary Table S5). To do a fair comparison, for every method, we computed LD scores and LDAK weights by using in-sample LD, and in all simulations we aim to estimate the SNP-heritability explained by the same set of M SNPs. Boxplot whiskers extend to the minimum and maximum estimates located within 1.5× interquartile range (IQR) from the first and third quartiles, respectively. Each box plot represents estimates from 100 simulations. Diamond points and error bars represent the mean and ± 2 SE centered on estimated heritability respectively. Mean and standard errors (SE's) are computed from 100 simulations.



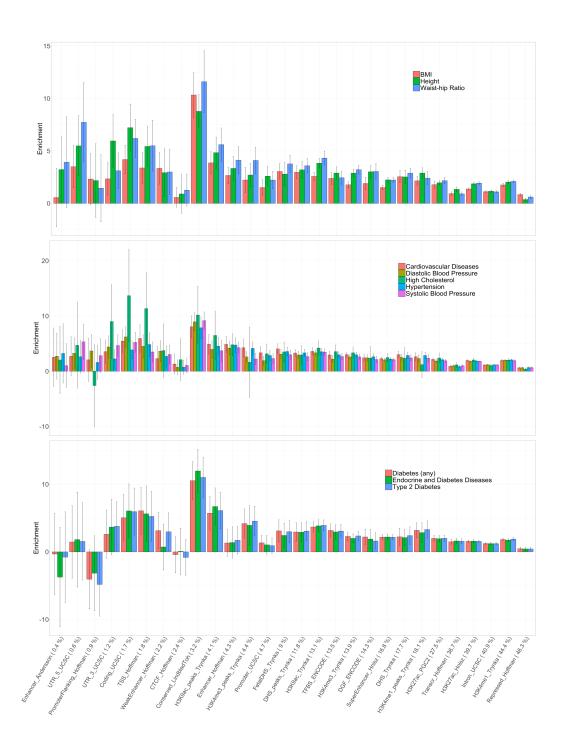
Supplementary Figure 5: Comparison of RHE-mc (red color) with GCTA-mc(blue color) in estimating partitioned heritability under 8 different genetic architectures on small-scale simulated data (M=590k array SNPs and N=10k individuals): We partition SNPs into 24 bins based on 6 MAF bins and 4 LDAK bins (Methods). True total SNP heritability is 0.25. Here x-axis represents the partitions (i.j denotes the bin defined based on i-th ldak bin and j-th MAF bin. The lower bin number denotes the lower MAF (LDAK weights). For example, bin 1.6 contains SNPs which are in the first quartile of LDAK weights and MAF> 0.05). y-axis represents the heritability. Each boxplot shows the distribution of estimates from 100 simulations. Note that GCTA-mc did not run successfully on all 100 simulations. Boxplot whiskers extend to the minimum and maximum estimates located within $1.5\times$ interquartile range (IQR) from the first and third quartiles, respectively. Each box plot represents estimates from 100 simulations. Diamond points and error bars represent the mean and ± 2 SE centered on estimated heritability respectively. Mean and standard errors (SE's) are computed from 100 replicates.

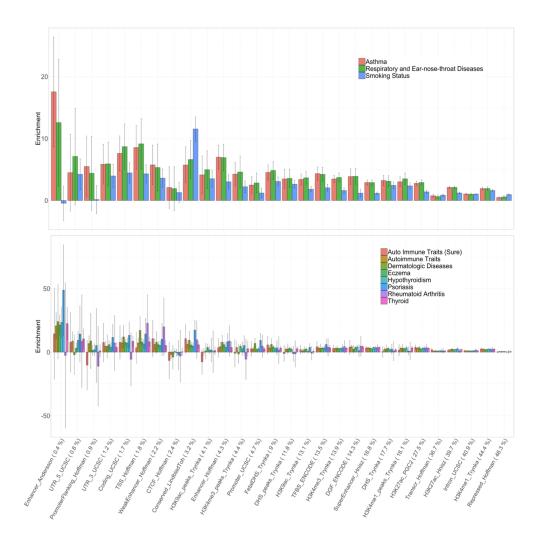


Supplementary Figure 6: Partitioned heritability estimates from RHE-mc on large-scale simulated data (M=590K array SNPs and N=337K individuals): We ran RHE-mc with 24 bins based on 6 MAF bins and 4 LDAK bins (Methods) over 8 different genetic architectures. Here x-axis represents the partitions (i.j denotes the bin defined based on i-th ldak bin and j-th MAF bin) and y-axis represents the heritability. Boxplot whiskers extend to the minimum and maximum estimates located within $1.5\times$ interquartile range (IQR) from the first and third quartiles, respectively. Each box plot represents estimates from 100 simulations. Diamond points and error bars represent the mean and ± 2 SE centered on estimated heritability respectively. Mean and standard errors (SE's) are computed from 100 replicates.

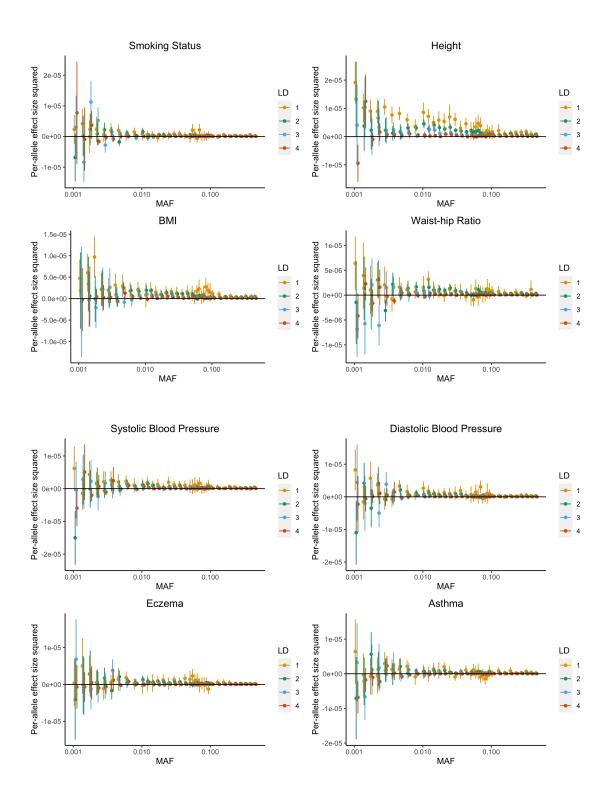


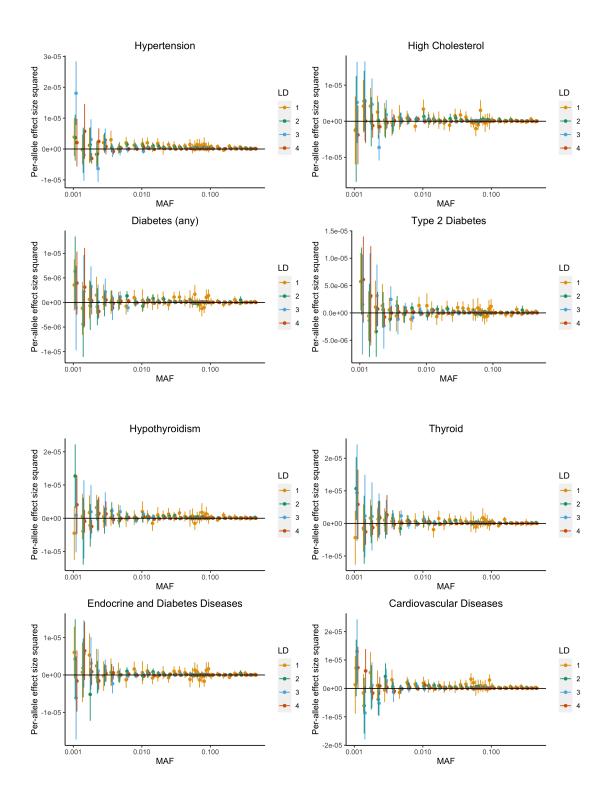
Supplementary Figure 7: Estimates of genome-wide SNP heritability from RHE-mc for 22 complex traits and diseases in the UK Biobank: We restricted our analysis to N=291,273 unrelated white British individuals. First, we applied RHE-mc to M=459,792 array SNPs (MAF> 1%) with 8 MAF/LD bins. Second, we applied RHE-mc to M=4,824,392 imputed SNPs (MAF> 1%) with 8 MAF/LD bins(Methods). Third, we applied RHE-mc to M=7,774,235 imputed SNPs (MAF> 0.1%) with 144 MAF/LD bins (Methods). Black bars mark ± 2 standard errors centered on estimated heritability.

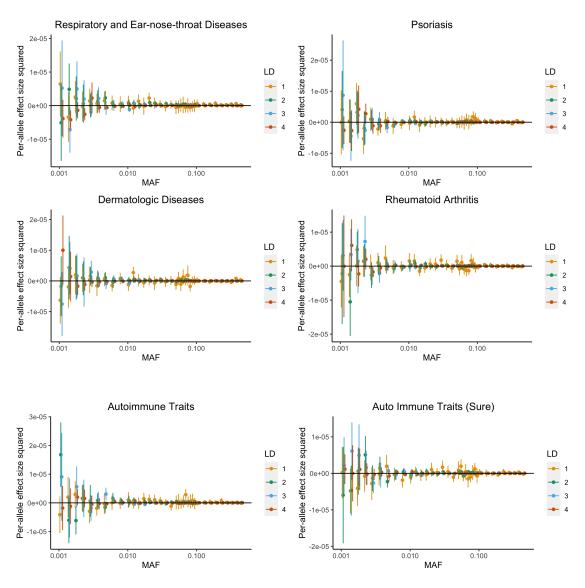




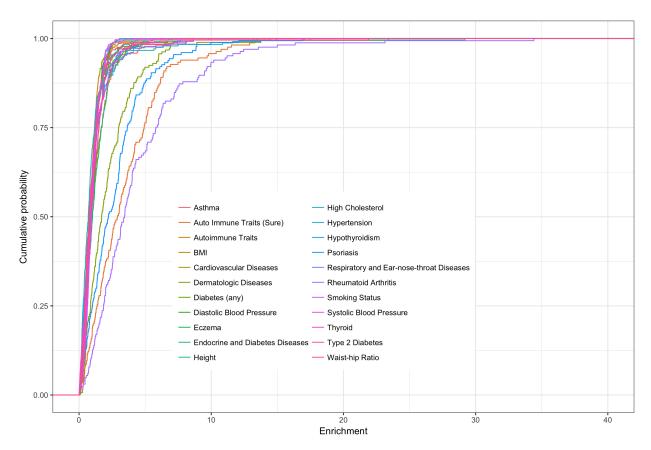
Supplementary Figure 8: Enrichment of heritability across 28 functional annotations: We applied RHE-mc to N=291,273 unrelated white British individuals and M=5,670,959 imputed SNPs (MAF >0.1% and present in 1000 Genomes Project). SNPs were partitioned based on 28 functional annotations that were defined in a previous study [6]. We grouped 22 traits in the UK Biobank into five categories (autoimmune, diabetes, respiratory, anthropometric, cardiovascular). Black bars mark ± 2 standard errors centered on estimated enrichment. Annotations are ordered by the proportions of SNPs in that annotation (given in parentheses)



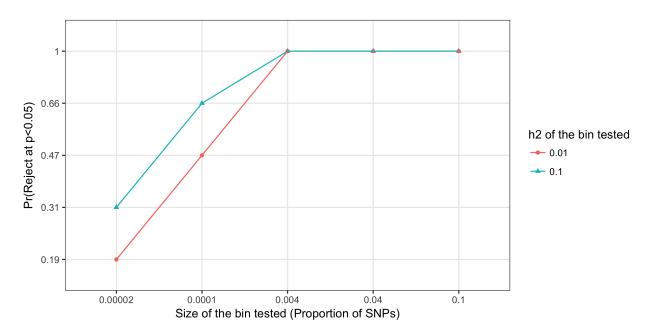




Supplementary Figure 9: **Per-allele effect size squared of 22 traits as a function of MAF**: We applied RHE-mc to N=291,273 unrelated white British individuals and M=7,774,235 imputed SNPs. SNPs were partitioned into 144 bins based on LD score (4 bins based on quartiles of the LD score with i denoting the i^{th} quartile) and MAF (36 MAF bins) (see Methods). Per allele heritability for bin k is defined as $\frac{h_k^2}{M_k*2f_k*(1-f_k)}$ where h_k^2 is the heritability attributed to bin k, M_k is the number of SNPs in bin k, and f_k is the average MAF in bin k. Points represent estimated per-allele heritability. Bars mark ± 2 standard errors centered on estimated per-allele heritability.



Supplementary Figure 10: Partitioning of genome-wide SNP heritability from RHE-mc for 22 complex traits and diseases in the UK Biobank (N=291,273 unrelated white British individuals, M=459,792 common SNPs with respect to 300 bins defined based on 10Mb base pairs. Here we plot the empirical cumulative probability respectively of the enrichment.



Supplementary Figure 11: Power as a function of annotation size. Each point represents a rejection probability over 100 simulations. All simulations have $h_{total}^2 = 0.7$, N = 291, 273, M = 459, 792, $p_{causal} = 0.05$.

3 Supplementary Tables

Number of random vectors	Point estimate (true SE)	Point estimate (SE of the estimate	
		due to randomization)	
10	0.24 (0.06)	0.24 (0.02)	
100	0.24 (0.05)	0.25 (0.001)	

Supplementary Table 1: Comparison of RHE-mc estimates with B=10 and B=100 on small scale (M=590K array SNPs and N=10k individuals). Here we quantify the contribution of randomization to the SE of the estimator. Here the true total heritability is 0.25. We first computed the SE of RHE-mc for B=10 and B=100 from 100 simulation replicates (second column). We then computed the SE of the estimates (due to the randomization) for a single replicate. For B=10, randomization contributes about a third of the total SE $(\frac{0.02}{0.06})$.

Genetic architecture				True SE	Jackknife SE
Percentage of	MAF of causal	MAF and LD	True h^2		
causal SNPs	SNPs	coupling			
0.01	[0.01, 0.05]	a=b=0	0.5	0.012	0.013
0.01	[0.01, 0.05]	a=0,b=1	0.5	0.018	0.015
0.01	[0.01, 0.05]	a=0.75,b=0	0.5	0.016	0.015
0.01	[0.01, 0.05]	a=0.75,b=1	0.5	0.013	0.013
0.01	[0.0,0.5]	a=b=0	0.5	0.009	0.013
0.01	[0.0,0.5]	a=0,b=1	0.5	0.016	0.014
0.01	[0.0,0.5]	a=0.75,b=0	0.5	0.018	0.019
0.01	[0.0,0.5]	a=0.75,b=1	0.5	0.012	0.015
0.01	[0.05, 0.5]	a=b=0	0.5	0.012	0.015
0.01	[0.05, 0.5]	a=0,b=1	0.5	0.021	0.017
0.01	[0.05, 0.5]	a=0.75,b=0	0.5	0.014	0.014
0.01	[0.05, 0.5]	a=0.75,b=1	0.5	0.015	0.017
1.0	[0.0,0.5]	a=b=0	0.5	0.007	0.007
1.0	[0.0,0.5]	a=0,b=1	0.5	0.007	0.007
1.0	[0.0,0.5]	a=0.75,b=0	0.5	0.006	0.006
1.0	[0.0,0.5]	a=0.75,b=1	0.5	0.007	0.008

Supplementary Table 2: Comparison of true SE with jackknife SE under 16 different genetic architectures: We defined 100 blocks over SNPs to estimate block jackknife SE. We ran RHE-mc with 24 bins based on 6 MAF bins and 4 LDAK score bins (see Methods). True SE is computed from 100 replicates for every setting. Jackknife SE yields estimates of true SE with relative bias -3% on average over 16 genetic architectures.

Genetic architecture		Heritability	y (GCTA-mc)	Heritability (RHE-mc)	
MAF of causal SNPs	MAF and LD coupling	Causal bin	Non-causal bin	Causal bin	Non-causal bin
[0.01, 0.05]	a = b = 0	0.244 ± 0.061	0.009 ± 0.051	0.242 ± 0.064	0.004 ± 0.052
[0.01, 0.05]	a = 0, b = 1	0.243 ± 0.062	0.008 ± 0.047	0.247 ± 0.060	0.003 ± 0.051
[0.01, 0.05]	a = 0.75, b = 0	0.241 ± 0.061	0.009 ± 0.050	0.240 ± 0.062	0.002 ± 0.051
[0.01, 0.05]	a = 0.75, b = 1	0.247 ± 0.056	$0.004 \pm\ 0.048$	0.244 ± 0.06	0.003 ± 0.051
[0.05, 0.5]	a=b=0	0.251 ± 0.048	0.012 ± 0.003	0.251 ± 0.052	0.007 ± 0.058
[0.05, 0.5]	a = 0, b = 1	0.248 ± 0.052	0.014 ± 0.054	0.240 ± 0.049	0.001 ± 0.055
[0.05, 0.5]	a = 0.75, b = 0	0.255 ± 0.047	0.000 ± 0.060	0.251 ± 0.052	0.000 ± 0.060
[0.05, 0.5]	a = 0.75, b = 1	0.250 ± 0.048	0.005 ± 0.05	0.241 ± 0.050	0.002 ± 0.058

Supplementary Table 3: Heritability contribution of causal bin vs non-causal bins on small-scale simulated data (M = 590k array SNPs and N = 10k individuals): We ran both RHE-mc and GCTA-mc with 24 bins based on 6 MAF bins and 4 LDAK bins for 8 genetic architectures (Methods). In all simulations, the proportion of causal variants is 0.01 and true total heritability is 0.25. The causal SNPs are restricted to lie within a specific range of MAF, *i.e.*, within [0.01, 0.05] for the first four rows and [0.05, 0.5] for the last four. Non-causal bins refer to those bins where none of the SNPs is causal, *i.e.*, in each of the first four genetic architectures, these would correspond to bins with MAF \notin [0.01, 0.05]. Causal bins refer to all remaining bins. Standard errors are computed from 100 replicates.

Genetic architecture				Heritability		
Percentage of	MAF of causal	MAF and LD	True total h^2	Causal bin	Non-causal bin	
causal SNPs	SNPs	coupling				
0.01	[0.01, 0.05]	a=b=0	0.5	0.501 ± 0.006	0.000 ± 0.004	
0.01	[0.01, 0.05]	a=0,b=1	0.5	0.498 ± 0.007	0.000 ± 0.003	
0.01	[0.01, 0.05]	a=0.75,b=0	0.5	0.500 ± 0.008	0.002 ± 0.004	
0.01	[0.01, 0.05]	a=0.75,b=1	0.5	0.490 ± 0.007	0.001 ± 0.003	
0.01	[0.05, 0.5]	a=b=0	0.5	0.501 ± 0.036	-0.001 ± 0.030	
0.01	[0.05, 0.5]	a=0,b=1	0.5	0.487 ± 0.012	0.005 ± 0.005	
0.01	[0.05, 0.5]	a=0.75,b=0	0.5	0.508 ± 0.026	-0.005 ± 0.023	
0.01	[0.05, 0.5]	a=0.75,b=1	0.5	0.490 ± 0.009	0.000 ± 0.005	

Supplementary Table 4: Heritability contribution of causal vs non-causal bins on large-scale simulated data (M = 590K array SNPs and N = 337K individuals): We ran RHE-mc with 24 bins based on 6 MAF bins and 4 LDAK bins (Methods). Standard errors are computed from 100 replicates.

MAF bin	Range	Number of SNPs
1	[0, 0.0126)	59330
2	[0.0126, 0.020)	59330
3	[0.020, 0.029)	59330
4	[0.029, 0.0433)	59330
5	[0.043, 0.0658)	59330
6	[0.065, 0.106)	59330
7	[0.106, 0.170)	59330
8	[0.170, 0.260)	59330
9	[0.260, 0.373)	59330
10	[0.373, 0.5)	59330

Supplementary Table 5: MAF bins which are used in running S-LDSC over the large scale simulated data.

True enrichment	Proportion of SNPs	point estimate	SE	Pr(rejection at p< 0.05)
2	0.4%	2.06	0.4	100%
1	0.4%	1.02	0.14	100%
0	0.4%	0.0	0.02	0.5%
2	0.01%	2.18	1.07	30%

Supplementary Table 6: Power as a function of annotation size. SE, point estimate, and probability of rejections are computed from 100 replicates. All simulations have $h_{total}^2 = 0.7$, N = 291, 273, M = 459, 792, $p_{causal} = 0.05$.

Trait	Heritability			
	Chromosome	MAF/LD	$10 \mathrm{Mb}$	
Autoimmune Traits	0.064 ± 0.005	0.054 ± 0.006	0.070 ± 0.004	
Auto Immune Traits (Sure)	0.011 ± 0.002	0.023 ± 0.006	0.029 ± 0.001	
Dermatologic Diseases	0.020 ± 0.003	0.0172 ± 0.003	0.021 ± 0.001	
Psoriasis	0.017 ± 0.002	0.014 ± 0.005	0.022 ± 0.003	
Rheumatoid Arthritis	0.008 ± 0.002	0.008 ± 0.002	0.010 ± 0.003	
Eczema	0.124 ± 0.007	0.104 ± 0.005	0.13 ± 0.006	
Hypothyroidism	0.097 ± 0.008	0.081 ± 0.005	0.11 ± 0.007	
Thyroid	0.095 ± 0.009	0.081 ± 0.008	0.109 ± 0.008	
Diastolic Blood Pressure	0.170 ± 0.005	0.145 ± 0.004	0.173 ± 0.003	
Systolic Blood Pressure	0.172 ± 0.006	0.146 ± 0.004	0.171 ± 0.004	
Cardiovascular Diseases	0.165 ± 0.006	0.134 ± 0.005	0.17 ± 0.004	
Hypertension	0.179 ± 0.006	0.150 ± 0.005	0.183 ± 0.006	
High Cholesterol	0.099 ± 0.015	0.070 ± 0.008	0.102 ± 0.003	
Diabetes (any)	0.069 ± 0.004	0.058 ± 0.003	0.072 ± 0.003	
Endocrine and Diabetes Diseases	0.064 ± 0.004	0.053 ± 0.003	0.065 ± 0.003	
Type 2 Diabetes	0.068 ± 0.004	0.057 ± 0.003	0.069 ± 0.005	
BMI	0.330 ± 0.014	0.264 ± 0.007	0.328 ± 0.013	
Height	0.583 ± 0.026	0.492 ± 0.017	0.59 ± 0.021	
Waist-hip Ratio	0.196 ± 0.009	0.167 ± 0.007	0.2 ± 0.005	
Asthma	0.122 ± 0.009	0.101 ± 0.006	0.127 ± 0.007	
Smoking Status	0.130 ± 0.004	0.111 ± 0.003	0.132 ± 0.002	
Respiratory and Ear-nose-throat Diseases	0.086 ± 0.007	0.071 ± 0.004	0.091 ± 0.004	

Supplementary Table 7: Estimates of genome-wide SNP heritability from RHE-mc for 22 complex traits and diseases in the UK Biobank (N=291,273 unrelated white British individuals, M=459,792 common SNPs). We run RHE-mc with 8 bins defined based on two MAF bins (MAF ≤ 0.05 , MAF> 0.05) and quartiles of the LD-scores. Furthermore, we run RHE-mc with 22 bins defined based on chromosome number. On average, partitioning based on chromosome numbers leads 21% higher estimates of genome-wide SNP heritability for 22 traits than partitioning based on MAF and LD. For instance, it leads 18% and 13% higher estimates of heritability for height and BMI respectively. We also partitioned SNP based on 10 Mb genomic regions (300 variance components).

References

- 1. Hans R Kunsch. The jackknife and the bootstrap for general stationary observations. *The Annals of Statistics*, 17(3):1217–1241, 1989.
- 2. Brendan K Bulik-Sullivan, Po-Ru Loh, Hilary K Finucane, Stephan Ripke, Jian Yang, Nick Patterson, Mark J Daly, Alkes L Price, Benjamin M Neale, Schizophrenia Working Group of the Psychiatric Genomics Consortium, et al. Ld score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nature genetics*, 47(3):291, 2015.
- 3. Steven Gazal, Hilary K Finucane, Nicholas A Furlotte, Po-Ru Loh, Pier Francesco Palamara, Xuanyao Liu, Armin Schoech, Brendan Bulik-Sullivan, Benjamin M Neale, Alexander Gusev, et al. Linkage disequilibrium—dependent architecture of human complex traits shows action of negative selection. *Nature genetics*, 49(10):1421, 2017.
- 4. Doug Speed, Gibran Hemani, Michael R Johnson, and David J Balding. Improved heritability estimation from genome-wide snps. *The American Journal of Human Genetics*, 91(6):1011–1021, 2012.
- 5. Doug Speed and David Balding. Better estimation of snp heritability from summary statistics provides a new understanding of the genetic architecture of complex traits. *bioRxiv*, page 284976, 2018.
- Hilary K Finucane, Brendan Bulik-Sullivan, Alexander Gusev, Gosia Trynka, Yakir Reshef, Po-Ru Loh, Verneri Anttila, Han Xu, Chongzhi Zang, Kyle Farh, et al. Partitioning heritability by functional annotation using genome-wide association summary statistics. *Nature genetics*, 47(11):1228, 2015.